

# AnyScript™ Reverse Transcriptase, 200 U/μl

LOT: See product label

EXPIRY DATE: See product label

## ORDERING INFORMATION

CAT. NO.	SIZE	PACKAGE CONTENT
BR0401001	20000 U	100 μl AnyScript Reverse Transcriptase 1 ml 5X Reverse Transcriptase Buffer

## COMPONENT

## COMPOSITION

AnyScript Reverse Transcriptase	AnyScript Reverse Transcriptase, 200 U/μl, in storage buffer containing 50% (v/v) glycerol.
5X Reverse Transcriptase Buffer	Optimized 5X Reaction Buffer for AnyScript Reverse Transcriptase.

## STORAGE

-30°C to -10°C (until expiry date – see product label)

## FEATURES

- Increased thermostability
- Exceptional performance for longer templates
- Excellent cDNA yield

## APPLICATIONS

- First-strand cDNA synthesis
- cDNA library construction
- RT-PCR

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## DESCRIPTION

biotechrabbit™ AnyScript Reverse Transcriptase is an improved and genetically engineered MMuLV Reverse Transcriptase with reduced RNase H activity and increased thermostability. The enzyme is ultrapure and free of RNases and Nucleases known to spoil RT reactions; this makes AnyScript a perfect choice for first strand cDNA synthesis at higher temperature than MMuLV. AnyScript is active up to 55°C with high yield and full-length cDNA product. The enzyme is tailored to be employed in both two step and one step RT-PCR and qRT-PCR with high sensitivity and specificity.

Superb sensitivity and specificity make AnyScript an ideal enzyme for fast RT reaction from low template input in diagnostics application.

## PROTOCOL

### Prevention of cDNA synthesis reaction contamination

RNase contamination is an exceptional concern when working with RNA. RNase A, providing most threat to RNA integrity, is a highly stable contaminant of any laboratory. To prevent RNA from degradation and to minimize possibility of contamination during cDNA synthesis; follow the guidelines below:

- Use separate clean areas for preparation of the samples and the reaction mixture.
- DEPC-treat all tubes and pipette tips or use certified nuclease-free labware with aerosol filters.
- Wear fresh gloves when handling RNA and all reagents.
- Always assess the integrity of RNA prior to cDNA synthesis in denaturing agarose gel electrophoresis.
- Use RNase free water and other reagents.
- To prevent RNA from degradation, add Ribonuclease inhibitor (optional) in to the cDNA synthesis reaction (20 units for 20 μl reaction).

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## Typical cDNA synthesis reaction set up

- Thaw on ice and mix very well all reagents.
- Assemble and keep all reactions on ice.
- To use time and reagents effectively, always prepare master mix for multiple reactions. For a master mix volume, always calculate the number reactions that you need plus one additional.
- Combine the following in an RNase-free reaction tube:

COMPONENT	VOLUME	FINAL CONCENTRATION
dNTP Mix (10 mM each)	1 μl	0.5 mM (each dNTP)
RNase Inhibitor, 40 U/μl (optional)	0.5 μl	1 U/μl
<i>Oligo (dT)<sub>12-18</sub> (10 μM) – or</i>	<i>2 μl</i>	<i>1 μM</i>
<i>Hexamer Primer (100 μM) – or</i>	<i>2 μl</i>	<i>10 μM</i>
<i>Gene Specific Primer (10 μM)</i>	<i>0.2 - 2 μl</i>	<i>0.1 - 1 μM</i>
5X Reverse Transcriptase Buffer	4 μl	1×
RNA Template	50 ng – 2 μg total RNA or 50–500 ng mRNA (polyA)	
AnyScript Reverse Transcriptase, 200 U/μl	0.5 – 1 μl	10 – 20 U/μl
RNase-free water	Variable	
Total volume	20 μl	

- Mix and collect the drops by centrifuging briefly
- When using
  - Hexamer Primer, incubate 10 minutes at 30°C followed by 37–50°C for 30 minutes (increase up to 60 min, if needed)
  - Oligo (dT) or gene-specific Primer incubate at 37–50°C for 30 minutes (increase up to 60 min, if needed)
- Reaction temperature can be raised to 55°C (small activity reduction may occur)
- In case of higher secondary structures, perform a pre-incubation of the RNA template for 5 min at 65°C (place on ice immediately after) and/or increase the incubation time for the subsequent RT reaction (at 37–50°C)
- Inactivate enzyme at 95°C for 5 minutes.
- Collect the drops by spinning briefly
- Store products at –20°C or proceed to next step, like PCR or qPCR
- Use maximum 10 μl of the cDNA synthesis reaction mix for PCR in 50 μl volume

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## CERTIFICATE OF ANALYSIS

### Unit Definition

One unit is the amount of enzyme activity that incorporates 1 nmol of dTTP into acid insoluble fraction in 10 minutes at 42°C when poly(A)+ RNA and oligo(dT)20 are used as template–primer.

### Quality Control

#### Exonuclease Activity

Linearized lambda/HindIII DNA fragments are incubated with the enzyme in a 50 μl reaction mixture for 4 h at 37°C. No DNA degradation observed.

#### Endonuclease/Nick Activity

Supercoiled plasmid DNA is incubated with the enzyme in a 50 μl reaction mixture for 4 h at 37°C. No conversion of covalently closed circular DNA to nicked DNA detected.

#### Contamination with *E. coli* DNA

Absence of *E. coli* genomic DNA is confirmed by qPCR using a sample of the enzyme and specific primers targeting the *E. coli* 16S rRNA gene. No contamination detected.

#### RNase Assay

An RNA template is incubated with the enzyme in a 20 μl reaction mixture for 1 h at 42°C. No RNA degradation observed.

#### Functional Assay

cDNA synthesis with Oligo (dT) and/or Hexamer primers, followed by PCR.

Quality confirmed by: Head of Quality Control

## SAFETY INSTRUCTIONS

For safety instructions please see Safety Data Sheets (SDS)/Sicherheitshinweise finden Sie in den SDS unter: <http://www.biotechrabbt.com/support/documentation.html>.

## USEFUL HINTS

- Visit Applications at [www.biotechrabbt.com](http://www.biotechrabbt.com) for more products and product selection guides.
- Most biotechrabbt products are available in custom formulations and bulk amounts.

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